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Waterpipe Smoke Effect on Oxidative Stress Levels in Buccal Cells Using MAWI i-SWAB Tubes

Joelle Mouawad¹, Joelle Abi Azar² and Marcel Bassil^{2*}

- 1 Faculty of Science, Lebanese University, Hadath, Lebanon
2 Biotechnology Department, Benta Pharma Industries, Dbayeh, Lebanon

*Corresponding author: Bassil M

✉ marcel.bassil@bpi.com.lb

Bassil M, Biotechnology Department, Benta Pharma Industries, Dbayeh, Lebanon,.

Tel: +9614545000

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Abstract

Background: Oxidative stress is an imbalance between reactive species and antioxidants production, leading to protein, DNA and lipid damage. The aim of this study was to evaluate the oxidative stress levels in buccal cells of ten non-shisha and heavy shisha smokers using MAWI iSWAB protein tubes.

Materials and findings: The presence of S-glutathionylation, NOX2 and iNOS was analyzed in buccal cells of ten non-smokers and ten heavy shisha smokers using MAWI i-SWAB protein tubes, knowing that they were able to detect previously several protein oxidation biomarkers (S-nitrosylation and nitrotyrosine). Proteins concentrations were assessed by Bradford assay then western blot method was used to analyze the presence of the different proteins of interest.

Conclusion: Results confirmed the findings about the ability of these tubes to detect S-nitrosylation, nitrotyrosine, as well as S-glutathionylation. Results showed also the presence of immature NOX2 in human buccal cells equally. Levels of iNOS were the same in non-shisha and heavy shisha smokers.

Keywords: Oxidative stress; Buccal cells; Shisha; S-glutathionylation; NOX2; iNOS; MAWI iSWAB tubes

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Introduction

Oxidative stress is the disruption of the redox signaling due to an overproduction of reactive species, or an impairment of the antioxidant defense system resulting in the pathogenesis of many diseases such as cancer, cardiovascular, pulmonary and neurodegenerative diseases as well as aging processes [1,2]. The reactive species, divided into free and non-radicals, are classified into three groups: Reactive Oxygen Species (ROS), Reactive Nitrogen Species (RNS), and Reactive Chlorine Species (RCS) [3].

One of the reactive oxygen species, superoxide anion, is produced enzymatically by nicotinamide adenosine dinucleotide phosphate oxidase (NADPH oxidase) [2]. NADPH oxidase 2 (NOX2) or gp91phox is one of the family members of NADPH oxidase. It's a membrane - embedded glycoprotein that needs p22phox protein for its stabilization. These proteins are inactive until they interact with other cytosolic components: p47phox, p67phox, p40phox and Rac. The activation consists of the phosphorylation of p47phox that enables the binding to p22phox and membrane lipids. It binds also to p67phox, promoting its interaction with

gp91phox. For a better association of the subunits and the main constituents, p40phox binds to p67phox as well. In resting cells, Rac is maintained in an inactive cytoplasmic complex with guanine nucleotide dissociation inhibitor (GDI). Upon activation, they dissociate and Rac is translocated to the membrane, following a GDP to GTP exchange promoted by a guanine nucleotide exchange factors (GEFs) [4].

Also, hydrogen peroxide is one of the reactive oxygen species that has the ability of oxidizing specific thiols of cysteine residues within proteins in a basic environment. One result of the formation of cysteine-sulfenic acid, is the high reactivity with the glutathione sulfhydryl (GSH) resulting in the formation of protein S-glutathionylation [5] known as biomarkers of exposure to hydrogen peroxide [6].

Nitric oxide is a free radical produced by nitric oxide synthases (NOS) when converting L-Arginine to L-Citrulline with NADPH and oxygen as cofactors. NOS has three isoforms: endothelial NOS (eNOS), neuronal NOS (nNOS) and inducible NOS (iNOS). The constitutive calcium-dependent NOS: eNOS is present in

the vascular endothelium and activated by arterial pressure changes, electric stimulation and several vasoactive substances such as bradykinin and angiotensin II; nNOS can be found in the postsynaptic terminals of the neuron [7]. The calcium-independent iNOS is induced by interleukins, cytokines and inflammations. Thus, if expressed, iNOS is active [8].

Once produced, NO could act directly or indirectly. The direct effect is done by activating guanyl cyclase (GC) (which produces GMPc and activates PKG) and promoting smooth muscles relaxation. The direct action of NO could also be on sulfhydryl of cysteine residues in proteins leading to S-nitrosylation, used as footprint of NO-mediated nitrosative stress [9,10]. The indirect roles are mainly by producing peroxynitrite through a reaction between NO and superoxide anion [2].

Peroxynitrite is a potent oxidant that can oxidize and destruct host cellular constituents (proteins, lipids, DNA), leading to the dysfunction of critical cellular processes and cell signaling pathways (**Figure 1**) [2]. The most predominant reaction of peroxynitrite is with tyrosine (known as nitration) and generates 3-nitrotyrosine. Due to peroxynitrite's high affinity for tyrosine residues in proteins, 3-nitrotyrosine has been used as a protein oxidation biomarker of peroxynitrite [9].

Oxidative stress has many endogenous and exogenous sources. One of the last, smoking or tobacco usage in all its form can cause oxidative stress. Water pipe, also known as shisha, involves the passage of charcoal-heated air through perforated aluminum foil which covers the tobacco placed in the head in order to generate smoke [11].

Waterpipe smoke is harmful. It causes impaired cellular growth and repair, inflammation, impaired vasodilation and oxidative stress [12]. The formation of free radicals, superoxide or nitric oxide result in lung oxidant toxicity through subsequent chain reactions resulting in uncontrolled destructive oxidation [13]. Hence, the damaging effects on cell function in lung epithelial cells and vascular endothelial cells are the two main factors in the development of chronic obstructive pulmonary disease (COPD), which is one of many respiratory diseases caused by tobacco smoke. Furthermore, oxidative stress and all these factors lead to cardiovascular diseases, including severe stenosis and ischemic heart disease. It affects also the heart rate variability, predictor of coronary artery disease. Besides, long history of waterpipe smoking increases the risk of having cardiovascular diseases [14].

Waterpipe smoking can also be a factor that increases the chance of having cancer due to genotoxic and clastogenic components in the water pipe smoke such as tar and polycyclic aromatic hydrocarbons [15].

However, the water pipe smoke effect on buccal cells has not been investigated yet, knowing that this type of cells represent an accessible pool of epithelial cells constituting a high diagnostic tool to evaluate the oxidative stress levels [16]. Besides, buccal cells collection is noninvasive and could be useful to provide information on host DNA, mRNA and proteins. In fact, these methods could be less time-consuming, and avoid the many limiting factors that could be faced such as inaccessibility and insufficiency of tissues. In addition, this collection method is

practical compared to other methods since it does not need special expertise. Thus, buccal cells collection could be useful to investigate the molecular and cellular basis of many diseases [17].

The MAWI i-SWAB tubes Protein kit is a new buccal cells collection methods that allows the collection and stabilization of proteins from the point of collection. They enable the mechanistic release of buccal cells in a specific lysis and stabilization buffer. These tubes are suitable for any population and for any age. They allow self-collection, less time-consuming and facilitates sample concentration, transportation, extraction, and long term storage at room temperature [18].

The aim of our study was to evaluate the effect of water pipe smoke on oxidative stress levels in buccal cells, using MAWI i-SWAB tubes for the detection of different protein oxidation biomarkers (S-glutathionylation, S-nitrosylation and tyrosine nitration). Furthermore, these tubes were used for the revelation of iNOS and NOX2.

Material and Methods

Study population

Buccal cells samples were collected from ten non-shisha and heavy shisha smokers, aged between 20 and 50 years. Participants were also asked to fill a questionnaire after signing the informed consent, which provided us with more information about their age, work, health status, shisha or cigarette smoking and more.

Sample collection

Members were asked to rinse their mouth twice with water and wait thirty minutes before applying swabbing. They had to rotate a cotton swab twenty times on both cheeks, and repeat the action twice. Swabs were then released into a 1 mL iSWABTM Protein (iSWAB-P-1200) tube for sample collection.

Sample processing

Samples were kept overnight at room temperature before extraction. They were centrifuged at 50.3 g for one minute at 23°C. Supernatants were then removed for protein dosage by Bradford Assay (following Sigma Aldrich protocol, Catalog Number B6916). After the preparation of a standard curve starting from [BSA]=2 mg/mL, absorbance of the samples was read by spectrophotometer at 595 nm. Each tube was assayed in duplicates.

Laemmli reducing buffer (BioRAD) containing Tris-HCl, β -mercaptoethanol, SDS, glycerol, and bromophenol blue was then added to the samples. Then samples were boiled for three minutes before performing western blot.

Western blot

After three minutes of boiling, 15 μ g of each sample were loaded in the 12% polyacrylamide gel's wells, and then run at 80-160 volts for two hours. Afterwards, gels were transferred to a nitrocellulose membrane (BioRad). Next, the membranes were stained with Ponceau Red for twenty minutes and then visualized by versa Doc imaging system (Bio-RAD).

Later, nitrocellulose membranes were blocked with 5% BSA (affymetrix) and incubated at 4°C overnight.

The day after, membranes were washed three times for five minutes with PBS 0.05% Tween 20 and incubated for 2 hours with primary antibody with gentle shaking at room temperature. Primary antibodies were: mouse monoclonal anti-nitrotyrosine (Abcam ab7048), mouse monoclonal anti-S-nitrosocysteine (Abcam ab94930), rabbit polyclonal anti-iNOS (Cell Signaling 2982), mouse monoclonal anti-glutathione (Abcam ab19534), rabbit polyclonal anti-NOX2 (Abcam ab31092) and anti-GAPDH.

Anti-glutathione gels were performed under non reducing conditions (Laemmli buffer without β-mercaptoethanol and boiling) and were then blocked with 5% milk and not BSA.

Anti-iNOS membranes were incubated overnight with the diluted antibody in 5% BSA, 1X PBS and 0.1% tween-20 at 4°C overnight.

After incubation with primary antibody, membranes were then washed three times with PBST for five minutes and incubated again with secondary antibodies for one hour at room temperature. The secondary antibodies were: HRP-conjugated rabbit anti-mouse polyclonal antibody (Abcam ab97046) and HRP-conjugated goat anti-rabbit antibodies.

TMB (Sigma Aldrich) was added to the membranes to be then visualized by Versa DOC imaging system.

Molecular weights of the bands obtained from the samples were compared to a broad range MW ladder (BioRAD, 161-0317) and to a pre-stained protein ladder-broad molecular weight (ab116028).

Results

The presence of the different protein oxidation biomarkers in non-smokers and smokers samples were detected by western blots.

In **Figure 2** bands of molecular weights ranging mainly between 30 KDa and 60 KDa revealed the presence of tyrosine nitration in buccal cells of non - shisha and heavy shisha smokers.

In **Figure 3**, bands of molecular weights mainly between 30 KDa and 50 KDa revealed the presence of S-nitrosylated proteins in buccal cells in non-smokers and heavy shisha smokers samples.

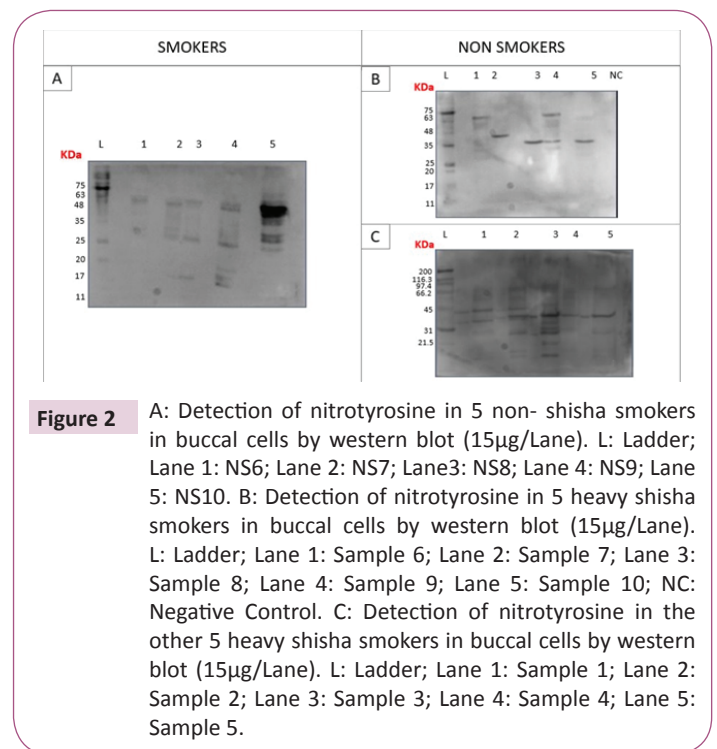
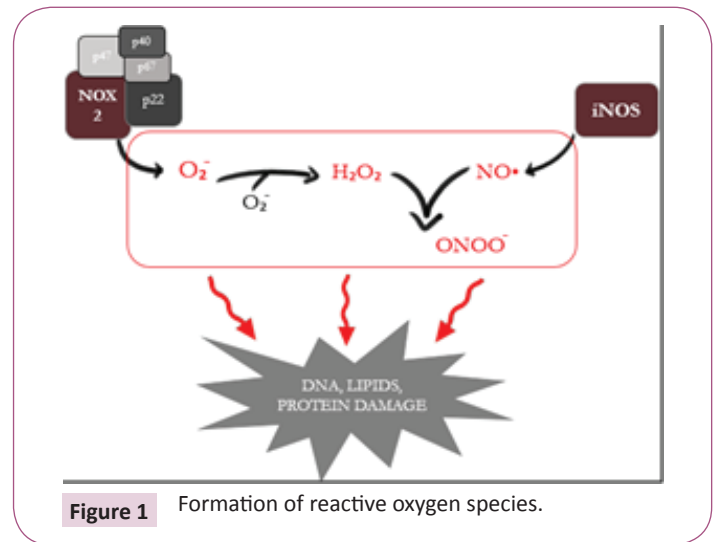
In **Figure 4**, bands at variable molecular weights revealed the presence of S-glutathionylated proteins in all samples.

In **Figure 5**, the anti-gp91phox antibody recognized proteins at 55 KDa, and bands had almost same intensities in all non-shisha and heavy shisha smokers samples.

Figure 6 shows the GAPDH band at 37 KDa. GAPDH is one of the many housekeeping proteins. It was used as an internal control for Western blot analysis to normalize different proteins expressions [19] in order to compare the levels of protein oxidation biomarkers in non- smokers and heavy shisha smokers.

In **Figure 7**, the presence of iNOS at the level of 135 KDa was revealed in buccal cells of non-shisha and heavy shisha smokers which are almost equally expressed.

Figure 8 shows the bands of GAPDH of 37 KDa in non-shisha smokers and heavy shisha smokers samples.



Discussion

The overall difference between the intensity of the bands in **Figures 1 and 2** is clear and shows a higher amount of tyrosine nitration in shisha smokers (**Figure 1**) compared to non-shisha smokers (**Figure 2**). However, housekeeping gene quantification such as GAPDH will be needed as internal loading control for further experiments.

Concerning the S-nitrosylated proteins obtained, our finding correlates with the previous ones of Bassil et al about the ability of MAWI tubes to preserve the protein oxidation biomarkers of nitric oxide and peroxynitrite: S-nitrosylation and tyrosine nitration [20]. The variation of the bands intensities could be correlated to the lifestyle and health condition of the participant.

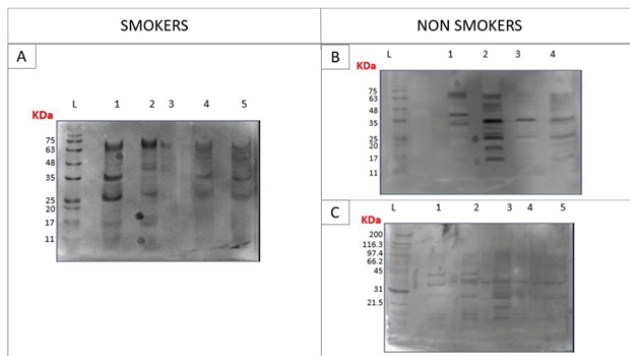


Figure 3 (A): Detection of S-nitrosylation in five non-shisha smokers in buccal cells by western blot (15µg/Lane). L: Ladder; Lane 1: Sample 6; Lane 2: Sample 7; Lane 3: Sample 8; Lane 4: Sample 9; Lane 5: Sample 10. (B): Detection of S-nitrosylation in 5 heavy shisha smokers in buccal cells by western blot (15µg/Lane). L: Ladder; Lane 1: Sample 6; Lane 2: Sample 7; Lane 3: Sample 8; Lane 4: Sample 9. (C): Detection of S-nitrosylation in the other 5 heavy shisha smokers in buccal cells by western blot (15µg/Lane). L: Ladder; Lane 1: Sample 1; Lane 2: Sample 2; Lane 3: Sample 3; Lane 4: Sample 4; Lane 5: Sample 5.

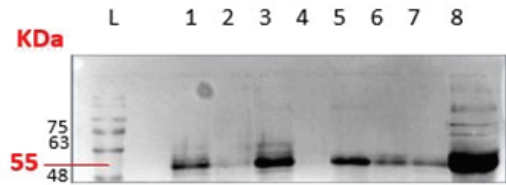


Figure 5 Detection of NOX2 in four heavy shisha smokers and four non-smokers samples in buccal cells by western blot (15µg/Lane). L: Ladder; Lane 1:S1; Lane 2:S2; Lane 3:S3; Lane 4:S4; Lane 5:NS1; Lane 6: NS2; Lane 7: NS3; Lane 8: NS4.

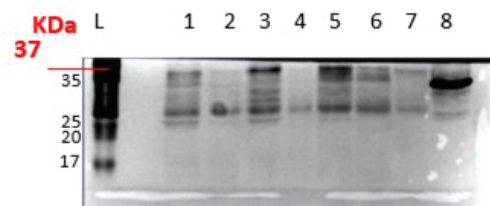


Figure 6 Detection of GAPDH in four heavy shisha smokers and four non-smokers in buccal cells by western blot (15µg/Lane). L: Ladder; Lane 1:S1; Lane 2:S2; Lane 3:S3; Lane 4:S4; Lane 5:NS1; Lane 6: NS2; Lane 7: NS3; Lane 8: NS4.

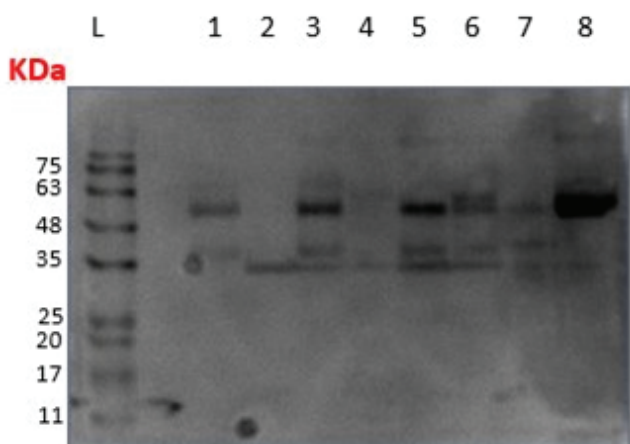


Figure 4 Detection of S-Glutathionylation in four heavy shisha smokers samples and four non-shisha smokers samples in buccal cells by western blot (15µg/Lane). L: Ladder; Lane 1:S1; Lane 2:S2; Lane 3:S3; Lane 4:S4; Lane 5:NS1; Lane 6: NS2; Lane 7: NS3; Lane 8: NS4..

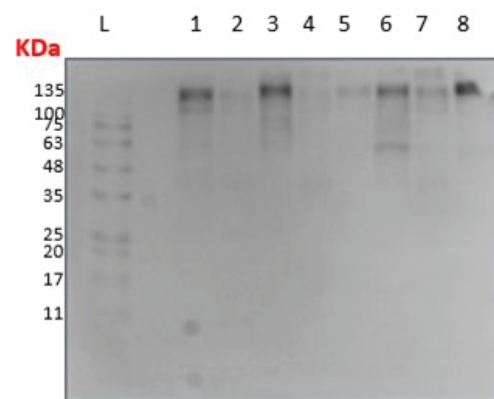


Figure 7 Detection of iNOS in four heavy shisha smoker samples and four non-shisha smokers samples in buccal cells by western blot (15µg/Lane). L: Ladder; Lane 1:S1; Lane 2:S2; Lane 3:S3; Lane 4:S4; Lane 5:NS1; Lane 6: NS2; Lane 7: NS3; Lane 8: NS4

It is important to note here that the appearance of oxidative stress biomarkers in non-shisha smokers is normal, because reactive oxygen species are always present. The damaging state is when these species overcome the antioxidant defense system [2].

The results of S-glutathionylation (**Figure 4**) show that the MAWI tubes can also be used for the detection of the protein oxidation biomarker of hydrogen peroxide in buccal cells: S-glutathionylation [6]. Thus, our results refer to the presence of hydrogen peroxide in both non-shisha and heavy shisha smoker samples. In addition to Bassil et al. MAWI protein tubes can also detect the presence of protein oxidation biomarker hydrogen

peroxide “S-glutathionylation” in human buccal epithelial cells [20].

The bands of 91 KDa represent the mature form of NOX 2 (glycosylated) and those of 55 KDa represents the immature form of NOX 2 (unglycosylated) [21]. These results show that NOX 2 in all samples are immature (**Figure 8**), which means that in all non-shisha and heavy shisha smokers samples, NOX 2 is expressed in its immature form.

The first step of NOX maturation is the glycosylation on arginine residues in the endoplasmic reticulum ER, before passing by the Golgi apparatus to finally attend the trans-membrane

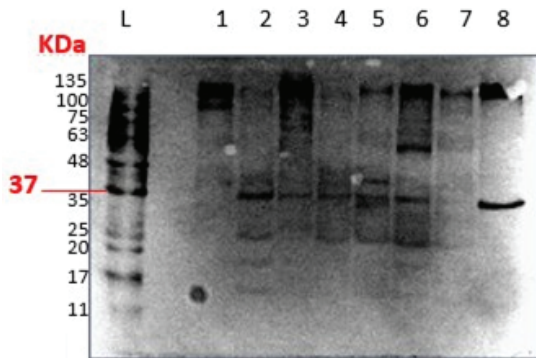


Figure 8 Detection of GAPDH in four heavy shisha smokers and four non- shisha smokers in buccal cells by western blot (15µg/Lane). L: Ladder; Lane 1:S1; Lane 2:S2; Lane 3:S3; Lane4:S4; Lane 5:NS1; Lane 6: NS2; Lane 7: NS3; Lane 8: NS4

[22]. Previous findings have shown that smoking and alcohol consumption induce cell death and ER stress in human lung epithelial cells and in pancreatic acinar cells [23]. ER stress induces dysfunction of the ER and disturbs its normal functions as folding, disulfide introduction and N-glycosylation and basically will not be able to glycosylate protein as NOX2 [24]. Based on that, these results could be correlated with our findings of unglycosylated NOX2 since all participants are alcohol consumers, heavy shisha smokers, and passive smokers.

In addition, our findings showed for the first time the presence of immature NOX2 in human buccal epithelial cells.

The comparison of iNOS expression **Figure 8** was unclear due to many factors. It could be because of the lifestyle of participants, since buccal cells are exposed to food and other agents that could make the comparison inaccurate, as well as patient's health status health status can highly affect the results [20]. Slomiany et al. findings about the increase of iNOS in buccal epithelial cells of alcohol consumers showed a positive correlation with our findings since most of the participants (non-shisha and heavy shisha smokers) were alcohol consumers [25].

Moreover, the small number of samples could also be a factor.

However, the groups of Hoyt have found that smoking decreases the expression of iNOS in lung epithelial cells by decreasing its mRNA transcription [26].

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The difference between our results and previous results could be due to the different localization of epithelial cells.

In addition, buccal epithelial cells have a higher proliferation rate than other keratinized cells [27]. That might also be a difference factor between results, since the exposure of smoking might not be optimal due to the high proliferation rate.

Conclusion

In conclusion, our results confirmed previous findings about the ability of MAWI tubes to detect the protein oxidation biomarkers S-nitrosylation and tyrosine nitration in buccal cell. As well as that, it showed that these tubes are able to detect S-glutathionylation (presence of hydrogen peroxide) found in buccal cells of the non-shisha and heavy shisha smokers. Our findings also included the presence of iNOS protein expression equally in non-shisha and heavy shisha smokers in epithelial buccal cells. Finally, our results showed for the first time the presence of the immature NOX2 in buccal epithelial cells equally in smokers and non-smokers and was also detectable by the same tubes.

For more significant results, our future research will include first of all the increase of our sample size. We will need also to confirm the immaturity of NOX2 by more advanced molecular biology tools in human epithelial buccal cells and the process behind this immaturity.

Author's Contribution

All authors participated in the analyses of the study and reviewed each drafts of the result article. MB and JM searched the databases and wrote the article. JAA contributed in experimental work. JM was responsible of sample collection.

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